Spectroscopic Evidence for the Redox Reaction between Copper(II) and a Pterin Cofactor Model, 6,7-Dimethyl-5,6,7,8-tetrahydropterin

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The reactions of Cu(II) with 6,7-dimethyl-5,6,7,8-tetrahydropterin dihydrochloride (H4DMP·2HCl) in CH3OH-CH3CN have been investigated by spectroscopic methods. The redox reaction in the 1:1 Cu(II)–H4DMP·2HCl system has been established by detection of the protonated trihydropterin radical (H4DMP•+) and its disproportionation products.

5,6,7,8-Tetrahydrobiopterin (1) is a natural pterin derivative required as the cofactor for the catalytic activities of phenylalanine hydroxylase (PAH) and other aromatic amino acid hydroxylases.<sup>1-3</sup>) In the presence of the cofactor PAH introduces a hydroxyl group into the benzene ring of phenylalanine by activating O2, converting phenylalanine to tyrosine in the biosynthesis of neurotransmitters, dopamine, norepinephrine, and epinephrine.<sup>1-3</sup>) The tetrameric mammalian PAH contains one non-heme iron per monomer, 1,2,4) whereas the monomeric PAH isolated from *Chromobacterium violaceum* <sup>5</sup>) contains a single type II copper<sup>6</sup>) with two coordinated histidyl imidazoles in the Cu(II) and Cu(I) states.<sup>7,8</sup>)

In the above hydroxylation reaction, 1 is oxidized to 7,8-dihydrobiopterin (quinonoid form).  $^{1,2}$ ) Although the oxidation may take place by a one-electron redox process via the trihydropterin radical, no evidence seems to have been reported on formation of the pterin radical species in the course of the enzymatic reaction. 6,7-Dimethyl-5,6,7,8-tetrahydropterin (H4DMP, 2), which also has the cofactor activity, has been shown to bind with the Cu(II) center of the bacterial PAH at the N(5) atom.  $^{9}$ ) A Cu(II) complex of tris(3-phenylpyrazolyl)hydroborate was observed to undergo a redox reaction with 5,6,7,8-tetrahydropterin (3) to give a Cu(I) complex in a preliminary experiment,  $^{10}$ ) but no radical species was reported. We have been studying the complex formation and redox reactions of pterin derivatives with Cu(II) complexes such as Cu(bpy) $^{2+}$  (bpy = 2,2'-bipyridine) $^{11}$ ) and established the structure of the first Cu(II) complex isolated.  $^{11}$ b) We here report the spectroscopic evidence for the redox reaction between Cu(II) and H4DMP·2HCl in CH3OH-CH3CN which gives Cu(I) and equal amounts of H4DMP and H2DMP (4) via a transient protonated trihydropterin radical H4DMP\*+.

Reduced pterins H<sub>2</sub>DMP·HCl·H<sub>2</sub>O and H<sub>4</sub>DMP·2HCl·H<sub>2</sub>O were prepared from 6,7-dimethylpterin<sup>12</sup>)

by reduction with Zn in 0.5 M KOH<sup>13</sup>) and catalytic hydrogenation with PtO<sub>2</sub> in 1M HCl (1 M = 1 mol dm<sup>-3</sup>), respectively. Rapid scan absorption spectra for the 1:1 [(C<sub>2</sub>H<sub>5</sub>)<sub>4</sub>N]<sub>2</sub>[CuCl<sub>4</sub>]–H<sub>4</sub>DMP·2HCl system in 50 v/v% CH<sub>3</sub>OH-CH<sub>3</sub>CN were measured at room temperature with a UNISOKU stopped flow–rapid scan spectrophotometer, the Cu(II) concentration being 0.5 mM. ESR spectra for the 1:1 Cu(NO<sub>3</sub>)<sub>2</sub>–H<sub>4</sub>DMP·2HCl system (2 mM) in 50 v/v% CH<sub>3</sub>OH-CH<sub>3</sub>CN or in 50 v/v% CD<sub>3</sub>OD-CD<sub>3</sub>CN with and without deuteration were obtained with a JEOL RE-1X ESR spectrometer at room temperature. The spectrum for the non-deuterated system was also measured at 77K by the rapid-mixing-freezing technique. <sup>1</sup>H NMR spectra (300 MHz) for the reaction mixture (10 mM) were measured in 50 v/v% CD<sub>3</sub>OD-CD<sub>3</sub>CN, after the ESR signals for H<sub>4</sub>DMP<sup>+</sup> had disappeared, with CD<sub>2</sub>HCN as the internal reference (δ 1.977 relative to TMS).

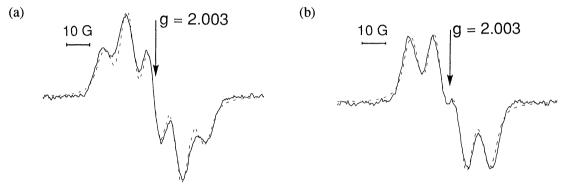


Fig. 1. Observed and simulated ESR spectra of 1:1  $Cu(NO_3)_2$ -H4DMP·2HCl in 50 v/v% CH3OH-CH3CN (2 mM)<sup>14</sup>) (a) and of the deuterated system in 50 v/v% CD3OD-CD3CN (b) under N2 at room temperature. Observed, ——; calcd., -----. Conditions: microwave frequency, 9.450 GHz; microwave power, 10 mW; modulation, 1G (1G = 0.1 mT).

Addition of Cu(NO3)2 in CH3CN (2 mM) to an equimolar amount of H4DMP·2HCl in CH3OH under N2 gave a colorless solution exhibiting the ESR spectrum due to a radical species at g = 2.003 at 298K (Fig. 1(a)), <sup>14)</sup> and no Cu(II) signals were observed. The spectrum is very similar to that reported for the pterin radical obtained by oxidation of 3 with H2O2 in CH3OH-CF3COOH. <sup>15,16)</sup> Deuteration of the exchangeable protons of H4DMP·2HCl prior to the reaction with Cu(NO3)2 gave the ESR spectrum of the pterin radical (Fig. 1(b)) with a hyperfine pattern different from that shown in Fig. 1(a). The spectra were reasonably simulated by the parameters <sup>17)</sup> analogous to those reported <sup>16b)</sup> and by considering the highest spin density on N(5) (0.358) with a lower density on N(8) (0.107) as indicated by EHMO calculations. The density on N(5) is supported by the previous calculations <sup>16b,18)</sup> and the transmethylation reactions of N-methylated tetrahydropterins. <sup>19)</sup> The ESR spectrum obtained for 1:1 Cu(NO3)2–H4DMP·2HCl by the rapid-mixing-freezing method showed both the unbound Cu(II) and radical signals, from which 40-50% of Cu(II) used was estimated to be reduced to Cu(I). The <sup>1</sup>H NMR spectrum of the final reaction mixture exhibited sharp, well-resolved methyl resonances of protonated H4DMP (6-CH3,  $\delta$  1.24 (d, J = 6.9Hz, 3H); 7-CH3,  $\delta$  1.29 (d, J = 6.6Hz, 3H)) and H2DMP (6-CH3,  $\delta$  2.38 (s, 3H); 7-CH3,  $\delta$  1.44 (d, J = 6.9Hz, 3H)), indicating that the tetra-and dihydro species were formed in approximately equal amounts and that there was no Cu(II) left unreacted.

Trihydropterins H<sub>3</sub>P• are known to disproportionate to tetra- and dihydropterins, H<sub>4</sub>P and H<sub>2</sub>P, respectively, and the constant K<sub>com</sub> for the following equilibrium (comproportionation, Eq. 1)

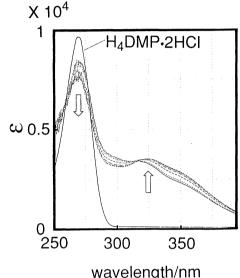
$$K_{com}$$

$$H_4P + H_2P \longrightarrow 2H_3P$$
 (1)

has been reported to be  $7.1 \times 10^{-7} \, \text{M}^{-1}$  for 6,6,7,7-tetramethyl-5,6,7,8-tetrahydropterin and its dihydro form in CF3COOH,<sup>20)</sup> which shows that the equilibrium lies far to the left, so that the amount of the radical species is very small after the equilibrium has been attained. The NMR spectrum is then interpreted as exhibiting the result of the disproportionation of H4DMP $^{\bullet+}$ . On the basis of the ESR signal intensity calibrated by that of 4-hydroxy-TEMPO (TEMPO = 2,2,6,6-tetramethylpiperidine-N-oxyl radical), the amount of the radical species in the present case was estimated to be 1.8% of H4DMP $^{\bullet}$ 2HCl used, and the K<sub>COM</sub> value was then calculated to be  $1.3 \times 10^{-3}$  according to Eq. 1.

Fig. 2. UV spectral changes for the 1:1  $[(C_2H_5)_4N]_2[C_4]-H_4DMP\cdot 2HCl$  system in the first 20 msec in 50 v/v% CH<sub>3</sub>OH-CH<sub>3</sub>CN.

Information on the transient species formed during the redox reactions was obtained from the absorption spectral changes in the UV and visible regions measured at room temperature with the stopped flow-rapid scan method. While the Cu(II) d-d band at 920 nm in the 1:1 [(C<sub>2</sub>H<sub>5</sub>)<sub>4</sub>N]<sub>2</sub>[CuCl<sub>4</sub>]- H<sub>4</sub>DMP·2HCl system in 50 v/v% CH<sub>3</sub>OH-CH<sub>3</sub>CN disappeared within 20 msec, the spectrum corresponding to the pterin moiety exhibited changes indi-



cating that the rapid oxidation of the tetrahydropterin in the first 5 sec proceeds in two steps, giving first an intermediate spectrum having a maximum at 270 nm and a broad peak at 325 nm with a shoulder at  $\sim$ 350 nm (Fig. 2) and then a spectrum with peaks at 270 and  $\sim$ 335 nm. Because H4DMP·2HCl has an absorption maximum at 268 nm and H2DMP·HCl has peaks at 254 and 361 nm in 50 v/v% CH3OH-CH3CN,<sup>21)</sup> the observed initial spectral changes may be interpreted as due to formation of the trihydropterin radical and its subsequent disproportionation.

The radical species of H4DMP obtained by oxidation with N3<sup>•</sup> has recently been reported to have a maximum at ~320 nm at pH 7,<sup>22</sup>) which corresponds well with the broad peak at 325 nm of the above intermediate spectrum. It is interesting to note in this connection that the oxidations of H4DMP by PAH,<sup>6</sup>) O2,<sup>18</sup>) and by the electrochemical method<sup>23</sup>) have been interpreted to indicate that the oxidation process involves one-electron steps with transient formation of pterin radicals. These and the present findings strongly suggest that in 1:1 Cu(II)-H4DMP·2HCl systems H4DMP undergoes a redox reaction with Cu(II) by a one-electron reaction step via formation of the trihydropterin radical, which then disproportionates to tetra- and dihydropterins.

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## References

- 1) S. Kaufman and D. B. Fisher, "Molecular Mechanisms of Oxygen Activation," ed by O. Hayaishi, Academic, New York(1974), p. 285.
- 2) R. L. Blakley and S. J. Benkovic, eds, "Folates and Pterins," Vol. 2, John Wiley & Sons, New York(1985); T. A. Dix and S. J. Benkovic, *Acc. Chem. Res.*, 21, 101-107(1988).
- 3) R. L. Blakley and V. M. Whitehead, "Folates and Pterins," Vol. 3, John Wiley & Sons, New York(1986); T. Nagatsu, S. Matsuura, and T. Sugimoto, *Med. Res. Rev.*, **9**, 25(1989).
- 4) D. B. Fisher, R. Kirkwood, and S. Kaufman, *J. Biol. Chem.*, **247**, 5161(1972); D. W. Gottschall, R. F. Dietrich, S. J. Benkovic, and R. Schiman, *ibid.*, **257**, 845(1982).
- 5) H. Nakata, T. Yamauchi, and H. Fujisawa, J. Biol. Chem., 254, 1829(1979).
- 6) S. O. Pember, J. J. Villafranca, and S. J. Benkovic, *Biochemistry*, 25, 6611(1986).
- 7) J. McCracken, S. O. Pember, S. J. Benkovic, J. J. Villafranca, R. J. Miller, and J. Peisach, *J. Am. Chem. Soc.*, **110**, 1069(1988).
- 8) N. J. Blackburn, R. W. Strange, R. T. Carr, and S. J. Benkovic, *Biochemistry*, 31, 5298(1992).
- 9) S. O. Pember, S. J. Benkovic, J. J. Villafranca, M. Pasenkiewicz-Gierula, and W. E. Antholine, *Biochemistry*, **26**, 4477(1987).
- 10) J. Perkinson, S. Brodie, K. Yoon, K. Mosny, P. J. Carroll, T. V. Morgan, and S. J. N. Burgmayer, *Inorg. Chem.*, 30, 719(1991).
- 11) a) T. Kohzuma, A. Odani, Y. Morita, M. Takani, and O. Yamauchi, *Inorg. Chem.*, **27**, 3854(1988); b) T. Kohzuma, H. Masuda, and O. Yamauchi, *J. Am. Chem. Soc.*, **111**, 3431(1989); c) A. Odani, H. Masuda, K. Inukai, and O. Yamauchi, *ibid.*, **114**, 6294(1992).
- 12) C. K. Cain, M. F. Mallette, and E. C. Taylor, Jr., J. Am. Chem. Soc., 68, 1996(1946).
- 13) W. Pfleiderer and W. Zondler, Chem. Ber., 99, 3008(1966).
- 14) O. Yamauchi, A. Odani, H. Masuda, and Y. Funahashi, "Bioinorganic Chemistry of Copper," ed by K. D. Karlin and Z. Tyeklár, Chapman & Hall, New York (1993), p. 363.
- 15) A. Ehrenberg, P. Hemmerich, F. Müller, T. Okada, and M. Viscontini, Helv. Chim. Acta, 50, 411(1967).
- 16) a) A. Bobst, Helv. Chim. Acta, **50**, 2222(1967); b) A. Bobst, ibid., **51**, 607(1968).
- 17) The following hyperfine coupling constants were used: For H4DMP $^{\bullet+}$ ,  $\alpha^{H(5)} = 8.0$ ,  $\alpha^{H(6)} = 8.9$ ,  $\alpha^{H(7)} = 2.0$ ,  $\alpha^{H(8)} = 2.0$ ,  $\alpha^{N(5)} = 7.0$ ,  $\alpha^{N(8)} = 2.0$  G, and line width = 2.5 G; for D2H2DMP·2HCl,  $\alpha^{D(5)} = 1.2$ ,  $\alpha^{H(6)} = 8.7$ ,  $\alpha^{H(7)} = 2.0$ ,  $\alpha^{D(8)} = 0.4$ ,  $\alpha^{N(5)} = 7.0$ ,  $\alpha^{N(8)} = 2.0$  G, and line width = 2.5 G.
- 18) J. A. Blair and A. J. Pearson, J. Chem. Soc., Perkin Trans. II, 1974, 80.
- 19) A. M. Bobst, Nature, 220, 164(1968).
- 20) G. Eberline, T. C. Bruice, R. A. Lazarus, R. Henrie, and S. J. Benkovic, *J. Am. Chem. Soc.*, **106**, 7916(1984).
- 21) The spectral data ( $\lambda_{max}/nm$  ( $\epsilon$ )) for H4DMP·2HCl and H2DMP·HCl measured in 50 v/v% CH3OH-CH3CN are as follows: H4DMP·2HCl, 268 (1.14 x 10<sup>4</sup>); H2DMP·HCl, 254 (1.65 x 10<sup>4</sup>), 361 (6.7 x 10<sup>3</sup>).
- 22) D. A. Armstrong, M. Farahani, and S. Surdhar, Can. J. Chem., 68, 1974(1990).
- 23) S. Kwee, Bioelectrochem. Bioenerg., 18, 79(1987).

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